

CML cells expressing IL-1RAP can be targeted by Chimeric Antigen Receptor–Engineered T Cells (CAR T-cells)

Mathieu Neto-Da-Rocha¹, Walid Warda¹, Rim Trad¹, Maxime Fredon¹, Sabeha Biichle¹, Fabrice Larosa², Etienne Daguidau², Francine Garnache-Ottou¹, Olivier Adotevi¹, Marina Deschamps¹ & Christophe Ferrand¹.

¹ INSERM UMR1098, EFS BFC, University of Bourgogne Franche-Comte, Besançon, France.
² Department of Hematology, University Hospital of Besançon, Besançon, France.



INTRODUCTION

In CML, gene expression profiling studies have revealed a cell-surface biomarker, **IL-1 receptor accessory protein (IL-1RAP, IL-1R3)**, that is expressed by the leukemic but not the normal CD34+/CD38- hematopoietic stem cell (HSC) compartment. Moreover, IL1RAP expression has been correlated with tumor burden and the clinical phase of CML. The tumor cell-surface expression makes IL1RAP an ideal candidate to target and eradicate CML HSCs, which are thought to be the origin of relapse. Thus, IL-1RAP is a promising cell-surface tumor-associated antigen for targeting with lower toxicity and higher efficacy in immunotherapy approaches, such as CAR T-cells. We hypothesized that T-cells expressing a CAR directed against IL1RAP may eliminate leukemic stem cells. Here, we describe the proof of concept and the preclinical evaluation of an IL1RAP CAR in the context of CML.

RESULTS I

1. Production of IL-1RAP CART-Cells.

The CAR lentiviral construct (pSDY-iC9-IL1RAPCAR-dCD19) was prepared by cloning the synthetically produced single-chain fragment variable (scFv; derived from the #A3C3 IL1RAP hybridoma, DiacloneSA, Besançon, FRANCE) into the SIN-pSDY backbone (Fig.1A). Lentiviral vector supernatant (SN) stock was harvested at 48 and 72 hours from subconfluent transfected 293T cells and overnight soft spin centrifugation. Activated T cells were established from healthy donors or patients and transduced with lentiviral SN encoding the IL-1RAP CAR or Mock sequence (missing the CAR sequence). The *in vitro* production process with lentiviral SN allowed for 82.4% to 78.4% transduction of primary T-cells (Fig.1B et C).

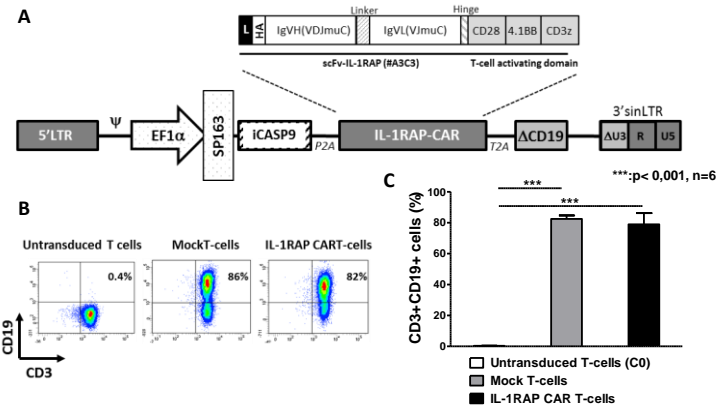


Figure 1 : Generation of IL1RAP CAR-expressing gene-modified T-cells.
A/ Lentiviral construct carrying the IL-1RAP CAR. **B/** Representative cytometry plot after CD3+/CD19+ staining of non transduced (C0), Mock-T, and IL-1RAP CAR T-cells. **C/** Results are presented as mean SD of 6 independent transductions of 6 different donor PBMCs (bottom right) using 100X concentrated SN.

RESULTS II

2. Inducible casp9 (iCASP9) safety.

In order to overcome potential toxicity of IL-1RAP CAR T-cells, we demonstrated the functionality of the **iCASP9/AP1903** (Rimiducid®) suicide system, (Fig.2A) included in our CAR construct (Fig.1A), *in-vitro* (Fig.2B) but also *in-vivo* in a NSG xenograft model (Fig.2C,D) that, when activate, is able to eliminate more than 90% of activated CART-cells, after 24hours of treatment with AP1903.

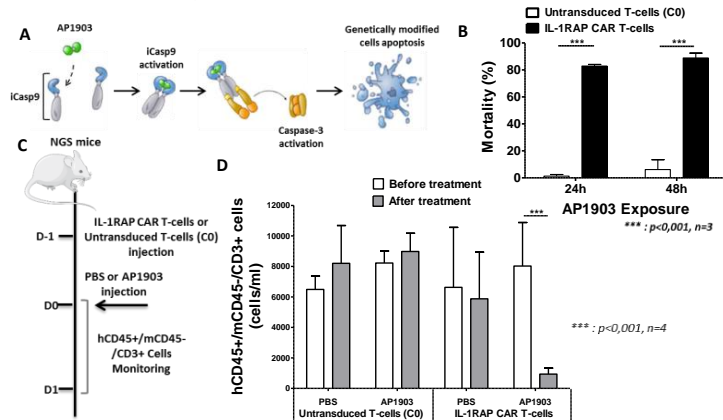


Figure 2 : iCASP9 safety.
A/ iCasp9 mechanism. **B/** Activation of iCASP9 *in vitro*. **C/** Activation of iCASP9 *in vivo* protocol. **D/** Activation of iCASP9 *in vivo*.

RESULTS III

3. Functionality of IL-1RAP CAR T-cells.

The proliferative capability of IL-1RAP CAR T-cells triggered by IL-1RAP target-expressing cells was determined by coculture (E:T 1:1) CFSE-stained (C0), Mock, or IL-1RAP CAR T-cells in the presence of K562, or KU812 cells. Effector IL-1RAP CAR T-cells divided significantly only in response to the presence of cell surface IL1RAP expressing and KU812 cells, and divided at a lowest levels against K562 (Fig.3A). IL1RAP dependent cytolytic potency of IL1RAP CAR expressing T-cells *in vitro* was determined using fluorescent (eFluor) and 7-AAD staining to discriminate CAR T-cells and living cells, respectively. As expected, coculture at an E:T ratio of 1:1 and compared with C0 or Mock T-cells revealed significant lytic activity characterized by the disappearance of cells in the 7-AAD-/eFluor- gate between IL-1RAP+ (KU812) target cells and IL-1RAP- (K562) target cells (Fig.3B). In a tumor xenograft murine model (Fig.3C), following tumor engraftment (D4), IL-1RAP CAR T-cells (E:T 1:1) were allowed to target CML KU812/Luc+ (i.v) tumors until a decrease in size. In contrast, we noticed tumor progression in untreated or Mock-T treated mice. Notably, tumors continued to grow in the absence of CAR T-cells in surviving mice in the untreated and Mock-T treated groups (Fig.3D).

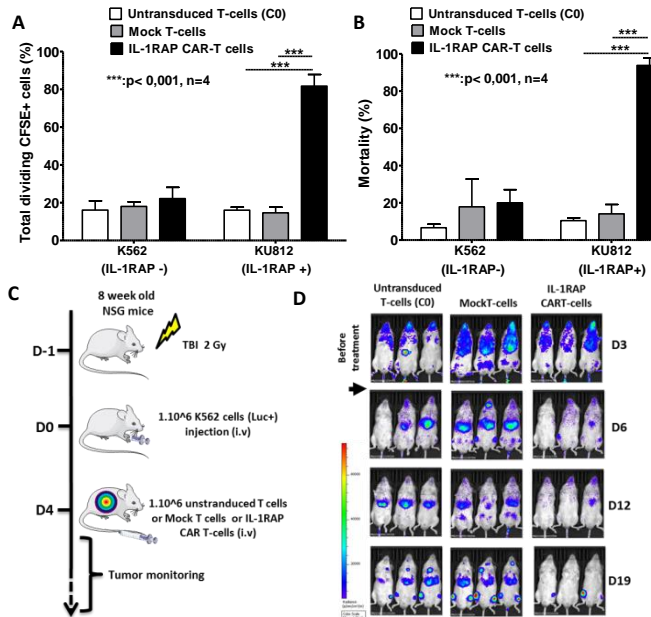


Figure 3 : Evaluation of proliferation and cytotoxicity of IL-1RAP CAR T-cells in vitro and in vivo.
A/ Non transduced (C0) and Mock or IL-1RAP CAR-transduced T-cells were cultured with k562 or KU812 at an ratio E:T of 1:1. Effectors were labeled with 0.5 mmol/L CFSE. After 48 measurement of CFSE dye dilution allowed assessment of the division of live CD3+/CD19+ gated cells. **B/** Effectors, labeled with e-Fluor, were cultured at an ratio E:T of 1:5 in the presence of IL-1RAP+ or - target cells and the percentage of total killed target cells is determined. **C/** Cytotoxicity *in vivo* protocol. **D/** *In vivo* tumor monitoring after xenograft (i.v) by KU812 and injection of C0, Mock T-cells or IL-1RAP CAR T-cells.

CONCLUSION

This preclinical work demonstrates for the first time the whole production and validation process of CAR T-cells directed against IL-1RAP-expressing CML stem cells, from the development and characterization of the mAb to the final *in vitro* and *in vivo* functional studies of gene-modified CAR T-cells. Taken together, these findings based on the CML model make IL-1RAP an interesting tumor-associated antigen for immunotherapy cell targeting using CAR T-cells. In-depth studies are required to determine and/or reduce potential toxicities and side effects before phase I clinical trials.

This work was the subject of a patent application and a publication in American journal for Cancer research (Warda et al., 2019)¹. This experimental approach may be applied in all CAR T-cells immunotherapies approaches developed at UMR1098.

¹Cancer Res, 2019 Feb 1;79(3):663-675. doi: 10.1158/0008-5472.CAN-18-1078.

