Murine IL-1α ELISA Kit

Instructions for use

Catalogue numbers: 1x96 tests: 660 010 096
2x96 tests: 660 010 192

For research use only

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Murine IL-1α ELISA Kit

1. Intended use

The murine IL-1α ELISA is an enzyme-linked immunosorbent assay for quantitative detection of murine Interleukin-1α (mIL-α) in cell culture supernatants, murine serum, plasma or other body fluids.

This kit has been configured for research use only. Not suitable for use in therapeutic procedures.

2. Introduction

2.1. Summary

The interleukin-1 (IL-1) species represent an important family of biologically active mono nuclear cell-derived proteins which are involved in inflammatory reactions and in immune responses (2, 8). Two distinct IL-1 species, IL-1α and IL-1β, have been identified (1, 6). They share similarities such as the same molecular weight, similar biological effects and the same receptors on target cells (4, 9). IL-1 proteins are produced by macrophages, monocytes and various other cell types such as adult T cell leukemias (10), fibroblasts, epithelial or endothelial cells (5), neutrophils and astrocytes (3). Their biological properties include pyrogenicity, bone resorption, presentation of antigen to T cells and stimulation of B and T lymphocyte proliferation (7).

IL-1α is an extracellular peptide, its activity has been demonstrated in various biological fluids (8).

2.2. Principle of the method

An anti-mIL-1α monoclonal coating antibody is adsorbed onto microwells.

mIL-1α present in the sample or standard binds to antibodies adsorbed to the microwells; a biotin-conjugated monoclonal anti-mIL-1α antibody is added and binds to mIL-1α captured by the first antibody.

Following incubation unbound biotin conjugated anti-mIL-1α is removed during a wash step. Streptavidin-HRP is added and binds to the biotin conjugated anti-mIL-1α. Following incubation unbound Streptavidin-HRP is removed during a wash step, and substrate solution reactive with HRP is added to the wells.

A coloured product is formed in proportion to the amount of mIL-1α present in the sample. The reaction is terminated by addition of acid and absorbance is measured at 450nm. A standard curve is prepared from six mIL-1α standard dilutions and mIL-1α sample concentration determined.

3. Reagents provided and reconstitution

REAGENTS (store at 2-8°C)	Quantity 1x96 well kit 660.010.096	Quantity 2x96 well kit 660.010.192	RECONSTITUTION
96-well precoated microtiter plate	1	2	Ready-to-use
Plate covers	2	4	
Biotin-Conjugate anti-mouse IL-1α monoclonal antibody	1 vial	2 vials	Dilute 100 times in Assay Buffer (70µl)
Streptavidin-HRP	1 vial	2 vials	Dilute 100 times in Assay Buffer (150µl)
mIL-1α Standard	2 vials	4 vials	Reconstitute as indicated on the vial
Assay Buffer Concentrate	1 vial	2 vials	(5 ml) 20X concentrate. Dilute in distilled water
Wash Buffer Concentrate	1 vial	2 vials	(50 ml) 20X concentrate. Dilute in distilled water
Sample Diluent	1 vial	2 vials	(12 ml)
Substrate Solution	1 vial	2 vials	(15 ml) Ready-to-use
Stop Solution (1M Phosphoric acid)	1 vial	2 vials	(15 ml) Ready-to-use

4. Materials required but not provided

- Microtiter plate reader fitted with appropriate filters (450nm required with optional 630nm reference filter)
- Microplate washer or wash bottle
- 10, 50, 100, 200 and 1,000µl adjustable single channel micropipettes with disposable tips
- 50-300µl multi-channel micropipette with disposable tips
- Multichannel micropipette reagent reservoirs
- Distilled water
- Vortex mixer
- Miscellaneous laboratory plastic and/or glass, if possible sterile

5. Storage Instructions

Store kit reagents between 2 and 8°C. Immediately after use remaining reagents should be returned to cold storage (2-8°C). Expiry of the kit and reagents is stated on box front labels.
The expiry of the kit components can only be guaranteed if the components are stored properly, and if, in case of repeated use of one component, the reagent is not contaminated by the first handling.

Wash buffer 1X: Once prepared, store at 2-8°C for up to 30 days.
Assay Buffer 1X: Once prepared, store at 2-8°C for up to 30 days.

6. Specimen collection, processing & storage

Cell culture supernatants, serum, plasma or other biological samples will be suitable for use in the assay. Remove serum from the clot or red cells, respectively, as soon as possible after clotting and separation.

Cell culture supernatants: Remove particulates and aggregates by spinning at approximately 1000 x g for 10 min.

Storage: If not analyzed shortly after collection, samples should be aliquoted (250-500µl) to avoid repeated freeze-thaw cycles and stored frozen at –70°C. Avoid multiple freeze-thaw cycles of frozen specimens.

Recommendation: Do not thaw by heating at 37°C or 56°C. Thaw at room temperature and make sure that sample is completely thawed and homogeneous before use. When possible avoid use of badly haemolysed or lipemic sera. If large amounts of particles are present these should be removed prior to use by centrifugation or filtration.

7. Safety & precautions for use

- Handling of reagents, serum or plasma specimens should be in accordance with local safety procedures , e.g.CDC/NIH Health manual : " Biosafety in Microbiological and Biomedical Laboratories" 1984
- Laboratory gloves should be worn at all times
- Avoid any skin contact with Stop Solution and TMB. In case of contact, wash thoroughly with water
- Do not eat, drink, smoke or apply cosmetics where kit reagents are used
- Do not pipette by mouth
- When not in use, kit components should be stored refrigerated or frozen as indicated on vials or bottles labels
- All reagents should be warmed to room temperature before use. Lyophilized standards should be discarded after use
- Once the desired number of strips has been removed, immediately reseal the bag to protect the remaining strips from deterioration
- Cover or cap all reagents when not in use
- Do not mix or interchange reagents between different lots
- Do not use reagents beyond the expiration date of the kit
- Use a clean disposable plastic pipette tip for each reagent, standard, or specimen addition in order to avoid cross contamination, for the dispensing of Stop Solution and substrate solution, avoid pipettes with metal parts
- Use a clean plastic container to prepare the washing solution
- Thoroughly mix the reagents and samples before use by agitation or swirling
- All residual washing liquid must be drained from the wells by efficient aspiration or by decantation followed by tapping the plate forcefully on absorbent paper. Never insert absorbent paper directly into the wells
- The TMB solution is light sensitive. Avoid prolonged exposure to light. Also, avoid contact of the TMB solution with metal to prevent colour development. Warning TMB is toxic avoid direct contact with hands. Dispose off properly
- If a dark blue colour develops within a few minutes after preparation, this indicates that the TMB solution has been contaminated and must be discarded. Read absorbance's within 1 hour after completion of the assay
- When pipetting reagents, maintain a consistent order of addition from well-to-well. This will ensure equal incubation times for all wells
- Follow incubation times described in the assay procedure
- Dispense the TMB solution within 15 min of the washing of the microtitre plate

8. Assay Preparation

Bring all reagents to room temperature before use

8.1. Assay Design

Determine the number of microwell strips required to test the desired number of samples plus appropriate number of wells needed for running zeros and standards. Each sample, standard and zero should be tested **in duplicate**. Remove sufficient Microwell Strips for testing from the aluminium pouch immediately prior to use. Return any wells not required for this assay with desiccant to the pouch. Seal tightly and return to 2-8°C storage.

Example plate layout (example shown for a 6 point standard curve)

	Standards		Sample Wells									
	1	2	3	4	5	6	7	8	9	10	11	12
A	500	500										
B	250	250										
C	125	125										
D	62.5	62.5										
E	31.25	31.25										
F	15.56	15.56										
G	0	0										
H												

All remaining empty wells can be used to test samples in duplicate

8.2. Preparation of Wash Buffer

If crystals have formed in the Wash Buffer Concentrate, warm it gently until they have completely dissolved.

Pour entire contents (50 ml) of the **Wash Buffer Concentrate** into a clean 1,000 ml graduated cylinder. Bring final volume to 1,000 ml with glass-distilled or deionized water. Mix gently to avoid foaming. The pH of the final solution should adjust to 7.4. Transfer to a clean wash bottle and store at 2°C to 8°C.

8.3. Preparation of Assay Buffer

Mix the contents of the bottle well.

Add contents of **Assay Buffer** Concentrate (5.0 ml) to 95 ml distilled or deionized water and mix gently to avoid foaming. Store at 2° to 8°C.

8.4. Preparation of Standard

Standard vials must be reconstituted with the volume of distilled water shown on the vial immediately prior to use. This reconstitution gives a stock solution of 1000pg/ml of mIL-1α. **Mix the reconstituted standard gently by inversion only**. Serial dilutions of the standard are made directly in the assay plate to provide the concentration range from 500 to 15.56 pg/ml. A fresh standard curve should be produced for each new assay.

- Add 100 µl of Sample Diluent to all standard and blank wells
- Immediately after reconstitution add 100µl of the reconstituted standard to well's A1 and A2, which provides the highest concentration standard at 500pg/ml. Mix the well contents by repeated aspirations and ejections taking care not to scratch the inner surface of the wells
- Transfer 100µl from wells A1 and A2 to B1 and B2. Mix the well contents by repeated aspirations and ejections taking care not to scratch the inner surface of the wells
- Continue this 1:1 dilution using 100µl from wells B1 and B2 through to wells F1 and F2 providing a serial diluted standard curve ranging from 500 to 15.56 pg/ml
- Discard 100µl from the final wells of the standard curve (F1 and F2)

Alternatively these dilutions can be performed in separate clean tubes and immediately transferred directly into the relevant wells.

8.5. Preparation of Biotin Conjugate

Make a 1:100 dilution with Assay Buffer (1X) in a clean plastic tube as needed according to the following table:

Number of Strips	Biotin-Conjugate (µl)	Assay Buffer (ml)
1 - 6	30	2.97
1 - 12	60	5.94

8.6. Preparation of Streptavidin-HRP

Make a 1:100 dilution of the concentrated **Streptavidin-HRP** solution in Assay Buffer (1X) as needed according to the following table:

Number of Strips	Streptavidin-HRP (µl)	Assay Buffer (ml)
1 - 6	60	6
1 - 12	120	12

9. Method

We strongly recommend that every vial is mixed thoroughly without foaming prior to use except the standard vial which must be mixed gently by inversion only.

Prepare all reagents as shown in section 8.
Note: final preparation of Biotin conjugate (section 8.5) and Streptavidin-HRP (section 8.6) should occur immediately before use.

Assay Step		Details
1.	Wash	Remove the pre-coated plate from the sealed pouch, removed any un-needed strips and wash the plate as follows: a) Dispense 0.3 ml of **1x washing solution** into each well b) Aspirate the contents of each well c) Repeat step a and b
2.	Addition	Prepare **Standard curve** as shown in section 8.4 above and add in duplicate to appropriate wells
3.	Addition	Add 100µl of **Sample Diluent** in duplicate to the blank wells
4.	Addition	Add 50µl of **Sample Diluent** to the sample wells
5.	Addition	Add 50µl of each **Sample** in duplicate to the designated wells
6.	Addition	Add 50µl of diluted **Biotin Conjugate** to all wells including blanks
7.	Incubation	Cover with a plastic plate cover and incubate at room temperature (18 to 25°C) for **2 hours** on a rotator set at 400rpm if available
8.	Wash	Remove the cover and wash the plate as follows: a) Aspirate the liquid from each well b) Dispense 0.3 ml of **1x washing solution** into each well c) Aspirate the contents of each well d) Repeat step b and c another two times
9.	Addition	Add 100µl of diluted **Streptavidin-HRP** solution into all wells
10.	Incubation	Cover with a plastic plate cover and incubate at room temperature (18 to 25°C) for **1 hour** on a rotator set at 400rpm if available
11.	Wash	Repeat wash step 8.
12.	Addition	Add 100µl of ready-to-use **TMB Substrate Solution** into all wells
13.	Incubation	Incubate for 1**0 minutes*** at room temperature on a rotar set at 200rpm if available. Avoid direct exposure to light by wrapping the plate in aluminium foil
14.	Addition	Add 100µl of **Stop Reagent** into all wells
Read the absorbance value of each well (immediately after step 14.) on a spectrophotometer using 450 nm as the primary wavelength and optionally 630 nm as the reference wave length (610 nm to 650 nm is acceptable).		

***Incubation time of the substrate solution is usually determined by the ELISA reader performance. Many ELISA readers only record absorbance up to 2.0 O.D. Therefore the colour development within individual microwells must be observed by the analyst, and the substrate reaction stopped before positive wells are no longer within recordable range*

10. Data Analysis

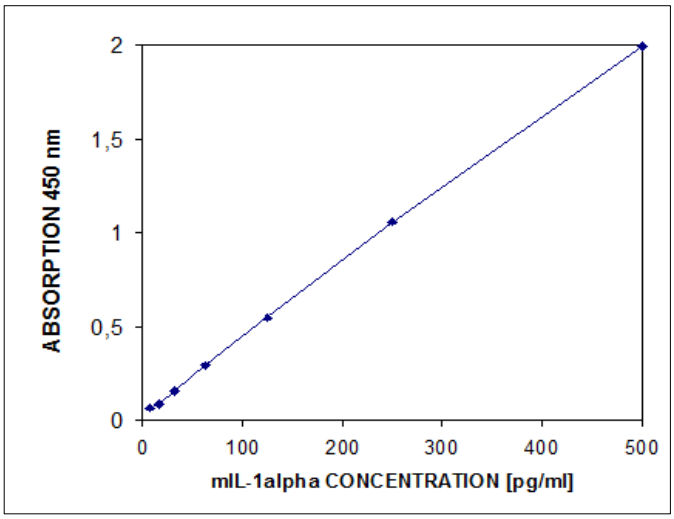
Calculate the average absorbance values for each set of duplicate standards and samples**.** Ideally duplicates should be within 20% of the mean.

Generate a linear standard curve by plotting the average absorbance of each standard on the vertical axis versus the corresponding mIL-1α standard concentration on the horizontal axis.

The amount of mIL-1α in each sample is determined by extrapolating OD values against mIL-1α standard concentrations using the standard curve**.**

Example mIL-1α Standard Curve

Standard	mIL-1α Conc	OD (450nm) mean	CV (%)
1	500	1.992	0
2	250	1.058	1.5
3	125	0.545	0.5
4	62.5	0.290	1.7
5	31.25	0.153	3.7
6	15.6	0.091	08
Zero	0	0.024	-



Note; curve shown above should not be used to determine results. Every laboratory must produce a standard curve for each set of microwell strips assayed.

For samples which have been diluted according to the protocol (1:2), the calculated concentration should be multiplied by the dilution factor.

11. Assay limitations

Do not extrapolate the standard curve beyond the maximum standard curve point. The dose-response is non-linear in this region and good accuracy is difficult to obtain. Concentrated samples above the maximum standard concentration must be diluted with Standard diluent or with your own sample buffer to produce an OD value within the range of the standard curve. Following analysis of such samples always multiply results by the appropriate dilution factor to produce actual final concentration.

The influence of various drugs on end results has not been investigated. Bacterial or fungal contamination and laboratory cross-contamination may also cause irregular results.

Improper or insufficient washing at any stage of the procedure will result in either false positive or false negative results. Completely empty wells before dispensing fresh Washing Buffer, fill with Washing Buffer as indicated for each wash cycle and do not allow wells to sit uncovered or dry for extended periods.

Disposable pipette tips, flasks or glassware are preferred, reusable glassware must be washed and thoroughly rinsed of all detergents before use.

As with most biological assays conditions may vary from assay to assay therefore **a fresh standard curve must be prepared and run for every assay**

12. Performance Characteristics

12.1. Sensitivity

The limit of detection of mIL-1α defined as the analyte concentration resulting in an absorption significantly higher than that of the dilution medium (mean plus two standard deviations) was determined to be 4.0 pg/ml (mean of 6 independent assays).

12.2. Specificity

The interference of circulating factors of the immune systems was evaluated by spiking these proteins at physiologically relevant concentrations into a mIL-1α positive serum. There was no cross reactivity detected.

12.3. Precision

Intra-assay

Reproducibility within the assay was evaluated in 3 independent experiments. Each assay was carried out with 6 replicates of 8 serum samples containing different concentrations of mouse IL-1α. 2 standard curves were run on each plate. **The overall intra-assay coeffiecient of variation has been calculated to be <5%.**

Inter-assay

Assay to assay reproducibility within one laboratory was evaluated in 3 independent experiments. Each assay was carried out with 6 replicates of 8 serum samples containing differents concentrations of mouse IL-1a. 2 standard curves were run on each plate. **The overall inter-assay coefficient of variation has been calculated to be <10%.**

12.4. Dilution Parallelism

4 serum samples with different levels of mIL-1α were assayed at serial two fold dilutions with 4 replicates each. Experiments showed an **overall mean recovery of 101 %.**

12.5. Spike Recovery

The spike recovery was evaluated by spiking four levels of mIL-1α into pooled normal murine serum. Recoveries were determined in two independent experiments with 4 replicates each. **The overall mean recovery was 75%.**

12.6. Stability

Storage Stability

Aliquots of spiked serum were stored at -20°C, 2-8°C, room temperature (RT) and at 37°C, and the mIL-1α level determined after 24 h. There was no significant loss of mIL-1α immunoreactivity during storage at -20°C, 4°C and room temperature. Storage at 37°C gave rise to about 30 % loss of mIL-1α immunoreactivity.

Freeze-thaw Stability

Aliquots of spiked serum were stored frozen at –20°C and thawed up to 5 times, and mIL-1α levels determined. There was no significant loss of IL-1α by freezing and thawing up to 5 times.

12.7. Expected values

There were no detectable mIL-1α levels found.
Elevated mIL-1α levels depend on the type of immunological disorder.

12.8. Standard Calibration

This immunoassay is calibrated against the International Reference Standard NIBSC 93/672. NIBSC 93/672 is quantitated in International Units (IU) and equivalence in ng/ml is indicated.
It has been calculated that 1 IU NIBSC corresponds to 1 pg mIL-1α.

13. Bibliography

1. Auron, P.E., Webb, A.C., Rosenwasser, L.J., Mucci, S.F., Rich, A., Wolff, S.M., and Dinarello, C.A. (1984). Nucleotide sequence of human monocyte interleukin 1 precursor cDNA. Proc. Natl. Acad. Sci. U.S.A. 81, 7907.

2. Dinarello, C.A. (1985). An update on human interleukin-1:from molecular biology to clinical relevance. J. Clin. Immunol. 5, 285.

3. Fontana, A., Kristensen, F., Dubs, R., Gemsa, D. and Weber, E. (1982). Production of prostaglandin E and an interleukin-1 like factor by cultured astrocytes and C6 glioma cells. J. Immunol. 129, 2413.

4. Kilian, P.L., Kaffka, K.L., Stern, A.S., Woehle, D., Benjamin, W.R., Dechiara, T.M., Gubler, U., Farrar, J.J., Mizel, S.B., and Lomedico, P.T. (1986). Interleukin-1* and interleukin-1* bind to the same receptor on T-cells. J. Immunol. 136, 4509.

5. Kurt-Jones, E.A., Fiers, W. and Pober, J.S. (1987). Membrane interleukin 1 induction on human endothelial cells and dermal fibroblasts. J. Immunol. 139, 2317.

6. March, C.J., Mosley, B., Larsen, A., Cerretti, D.P. Braedt, G., Price, V., Gillis, S., Henney, C.S., Kronheim S.R., Grabstein, K., Conlon, P.J., Hopp, T.P. and Cosman, D. (1985). Cloning sequence and expression of two distinct human interleukin-1 complementary DNAs. Nature 315, 641.

7. Mizel, S.B. (1987). Interleukin 1 and T-cell activation. Immunol. Today 8, 330.

8. Oppenheim, J.J., Kovacs, E.J., Matsushima, K., Durum, S.K. (1986). There is more than one interleukin1. Immunol. Today 9, 45.

9. Wood, D.D., Bayne, E.K., Goldring, M.B., Gowen, M., Hamerman, D., Humes, J.L., Ihrie, E.J., Lipsky, P.E. and Straruch, M.J. (1985). The four biochemically distinct species of human interleukin 1 all exhibit similar biologic activities. J. Immunol. 143, 895.

10. Yamashita, U., Shirakawa, F. and Nakamura, H. (1987). Production of interleukin 1 by adult T cell leukemia (ATL) cell lines. J. Immunol. 138, 3284.

14. Assay Summary

Total procedure length : 3h10mn

Add Sample Diluent, Sample , Standard and Biotin Conjugate
↓
Incubate 2 hours at room temperature
↓
Wash three times
↓
Add 100µl of Streptavidin-HRP
↓
Incubate 1 hour at room temperature
↓
Wash three times
↓
Add 100 µl of ready-to-use TMB
Protect from light. Let the color develop for 10 mn.
↓
Add 100 µl Stop Reagent
↓
Read Absorbance at 450 nm

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